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SPECTROSCOPIC PROPERTIES OF CHLOROPEROXIDASE COMPOUNDS II AND III — POSSIBLE STRUCTURAL MODELS FOR ANALOGOUS CYTOCHROME P-450 DERIVATIVES

Previous spectroscopic studies of analogous derivatives of chloroperoxidase (CPO) and cytochrome P-450 (P-450) have demonstrated that close similarities exist in their UV-visible absorption [1], magnetic circular dichroism (MCD) [2], EPR [3], Mössbauer [4], and extended X-ray absorption fine structure [5] properties. In particular, both enzymes exhibit unique hyperporphyrin («split Soret») spectra in their ferrous-CO [1] and ferric-thiolate [6] adducts, strongly suggesting the coordination of an endogenous thiolate ligand in CPO, as has well been established for P-450 [7]. These extensive parallels between the coordination properties of CPO and P-450 have motivated us to generate other CPO derivatives as potential analogs for catalytic intermediates of P-450, especially its oxygen complexes.

We report herein the generation of a unique, stable CPO-dioxygen adduct (CPO•O₂, CPO Compound III) and of CPO Compound II (CPO-II), and their characterization with UV-visible absorption and MCD spectroscopy. The CPO dioxygen adduct is formed by bubbling O₂ into a dithionite-reduced enzyme solution at -30°C in cryogenic mixed solvents. CPO-II is generated at

ambient temperature (~4°C) by adding peroxides and ascorbic acid to ferric CPO (1-2 μM) in a molar ratio of 100:2000:1. The UV-visible absorption and MCD spectra of CPO•O₂ and CPO-II are displayed in figs. 1 and 2, respectively.

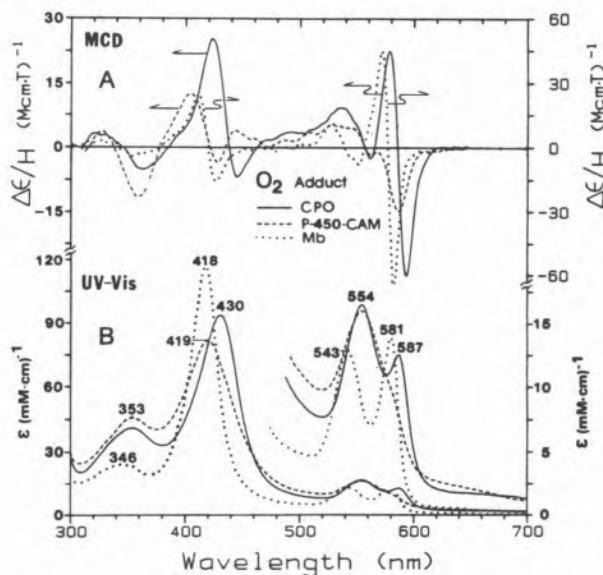


Fig. 1

UV-visible absorption (bottom) and MCD (top) spectra of oxygenated CPO (—), P-450-CAM (---) and Mb (·····). The spectra were obtained in 65% (v/v) ethylene glycol/0.035 M potassium phosphate buffer (pH 6.0, 7.4 and 7.0 before mixing, respectively) at -30°C with 30-40 μM protein concentrations. The P-450-CAM sample contained 100 mM KCl and 4 mM d-camphor

The absorption spectrum of CPO•O₂ at -30°C (Fig. 1B) [$\lambda_{nm}(\epsilon_{mM})$:354(41), 430(94), 554(16.5), 587(12.5)] has two noticeable distinctions from that of P-450•O₂: (a) the Soret peak of CPO•O₂ is red-shifted about 10 nm from that of P-450•O₂, and (b) a distinct α -peak is seen at 587 nm in the CPO case that is not observed for P-450•O₂. However, the overall spectral features of CPO•O₂ are more similar to those of P-450•O₂ [$\lambda_{nm}(\epsilon_{mM})$:353(46), 419(82), 554(16)] than of oxygenated myoglobin. Oxygenated CPO is EPR silent at 77 K. The bound O₂ in oxygenated CPO can be replaced by CO; upon bubbling CO into the CPO•O₂ solution at -30°C, the diagnostic hyperporphyrin spectrum of ferrous-CO CPO ($\lambda_{max,nm}$:362, 445, 549) is generated. CPO•O₂ undergoes autoxidation to form native ferric CPO without any detectable intermediates with a half life comparable to that of P-450•O₂. In fig. 2,

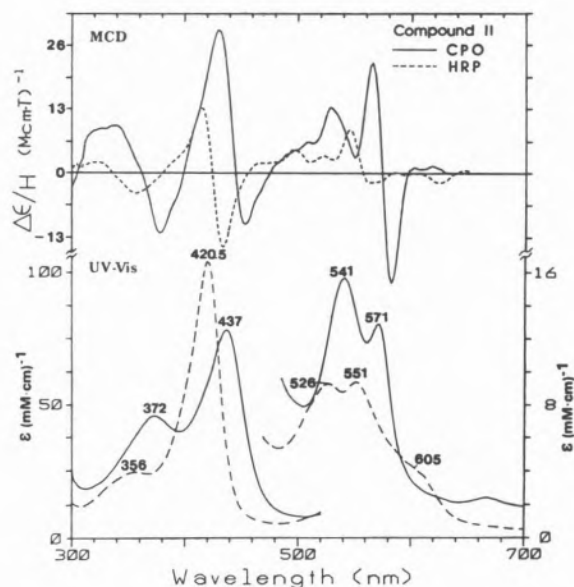


Fig. 2

UV-visible absorption (bottom) and MCD (top) spectra of Compound II derivatives of CPO (—) and HRP (----). The measurements for CPO (1.5–2 μ M) were made in 0.1 M potassium phosphate buffer, pH 6.0, with peroxides (~ 0.15 mM) and ascorbic acid (~ 3 mM) and those for HRP (10–15 μ M) in 0.005 M sodium carbonate buffer, pH 10.5, with equivalent amounts of EtOOH and ascorbic acid, at $\sim 4^\circ\text{C}$. The MCD spectrum of CPO Compound II is the average of 3 measurements

UV-visible absorption (bottom) and MCD (top) spectra of horseradish peroxidase Compound II (HRP-II) are overplotted with those of CPO-II. Considerable differences are seen between the spectra of HRP-II and CPO-II, presumably because the endogenous axial ligands are not the same: histidine imidazole for HRP and thiolate, most likely, for CPO.

In conclusion, the dioxygen adducts of CPO and P-450 are clearly distinguishable from that of Mb, a histidine-ligated heme protein, in their UV-visible absorption and MCD spectral properties [8]. Considerable spectral differences are also seen between the Compound II states of CPO and HRP. The small, but nonetheless significant, spectroscopic dissimilarities between the dioxygen adducts of CPO and P-450 (Fig. 1A, B) are somewhat surprising, since the analogous ferric and ferrous ligand adducts of CPO and P-450 show great similarities [1,6,9]. The reason for these differences are not clear at the present.

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REFERENCES

- [1] P.F. HOLLENBERG, L.P. HAGER, *J. Biol. Chem.*, **248**, 2630 (1973).
- [2] J.H. DAWSON, J.R. TRUDELL, G. BARTH, R.E. LINDER, E. BUNNENBERG, C. DJERASSI, R. CHIANG, L.P. HAGER, *J. Am. Chem. Soc.*, **98**, 3709 (1976).
- [3] P.F. HOLLENBERG, L.P. HAGER, W.E. BLUMBERG, J. PEISACH, *J. Biol. Chem.*, **255**, 4801 (1980).
- [4] P.M. CHAMPION, E. MÜNCK, P.G. DEBRUNNER, P.E. HOLLENBERG, L.P. HAGER, *Biochemistry*, **12**, 426 (1973).
- [5] S.P. CRAMER, J.H. DAWSON, K.O. HODGSON, L.P. HAGER, *J. Am. Chem. Soc.*, **100**, 7287 (1978).
- [6] M. SONO, J.H. DAWSON, L.P. HAGER, *J. Biol. Chem.*, **259**, 13209 (1984).
- [7] M. SONO, L.A. ANDERSSON, J.H. DAWSON, *J. Biol. Chem.*, **257**, 8308 (1982).
- [8] J.H. DAWSON, S.P. CRAMER, *FEBS Lett.*, **88**, 127 (1978).
- [9] J.H. DAWSON, M. SONO, L.P. HAGER, *Inorg. Chim. Acta*, **79**, 184 (1983).



PS1.42 — TH

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ANION BINDING TO A CYTOCHROME *c'*

The cytochromes *c'* are mono- and diheme proteins reported to comprise the largest and most widespread class of bacterial cytochromes known [1]. These proteins are similar to the low-spin